

Pearls & Oy-sters: SARS-CoV-2 Infection of the CNS in a Patient With Meningeosis Carcinomatosa

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Pearls

- Neurologic severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection can present with fever, headache, and meningism.
- For diagnosis of SARS-CoV-2 infection of the CNS, reverse transcription PCR (RT-PCR) from CSF samples should be performed, because viral RNA may be detected in CSF for longer periods than in respiratory samples.
- Determination of the SARS-CoV-2-specific immunoglobulin G (IgG) antibody index in CSF and serum is feasible for the detection of a SARS-CoV-2-specific intrathecal IgG antibody synthesis.

Oy-sters

- Neurologic SARS-CoV-2 disease can present without accessory respiratory symptoms and signs.
- Negative SARS-CoV-2 RT-PCR in respiratory specimens does not exclude SARS-CoV-2 infection of the CNS.
- In patients with meningeosis neoplastica presenting with acute neurologic symptoms, the possibility of concomitant CNS infection should always be considered.

Coronavirus disease 2019 (COVID-19), the disease caused by SARS-CoV-2, primarily manifests as respiratory illness. Disease severity ranges from asymptomatic infections to acute respiratory distress syndrome. Recent observations demonstrated that SARS-CoV-2 can also cause neurologic symptoms, which is known for many coronaviruses, including SARS-CoV-1 and Middle East respiratory syndrome coronavirus.¹ Risk factors for neurologic complications of COVID-19, clinical presentation, and the appropriate virologic diagnostic approach need to be further analyzed. We report a case of SARS-CoV-2 infection of the CNS in a patient with meningeosis carcinomatosa.

Case Presentation

A 53-year-old man was transferred to our hospital (day 1) with fever and increasing headache for 1 day. The day before admission, a nasopharyngeal swab was positive for SARS-CoV-2 RNA. The patient reported no respiratory symptoms. Due to suspected meningitis, an empirical anti-infective treatment consisting of ceftriaxone, ampicillin, and acyclovir was immediately initiated. Adenocarcinoma of the esophagus (ypT1bpN2M0R0) had been diagnosed 4 months earlier, which was resected after 4 cycles of neoadjuvant chemotherapy with the FLOT regimen (folinic acid, 5-fluorouracil, oxaliplatin, docetaxel). At neurologic examination, the patient presented with meningism and decreased vigilance. The patient did not show any signs

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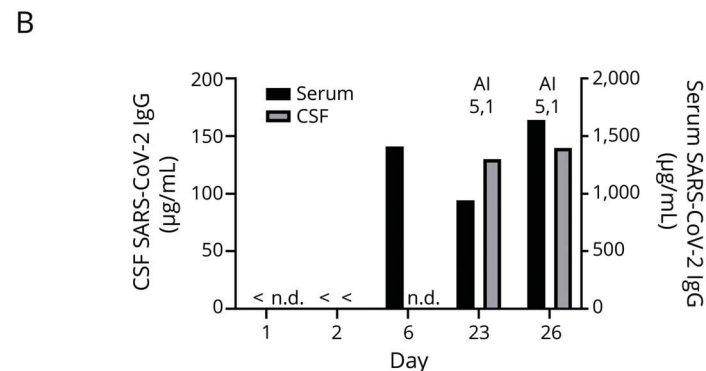
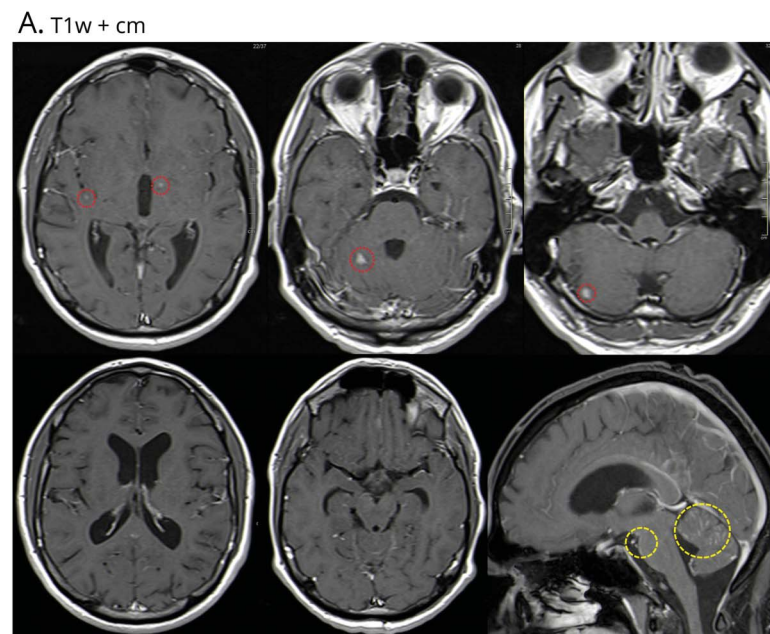
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of a respiratory infection. Respiratory rate was 12 breaths per minute and oxygen saturation was 96% without additional oxygen supply. Differential blood count showed leukocytopenia (2.88/nL) and normal neutrophil granulocyte count (2.46/nL) and absolute lymphocytopenia (0.34/nL). Together with normal C-reactive protein (2.8 mg/L), these laboratory findings were suggestive for a viral infection. CSF analysis on day 1 revealed lymphomonocytic pleocytosis (57 leukocytes/ μ L, 96.5% mononuclear cells; 3 red blood cells/ μ L) with elevated lactate (5.84 mmol/L) and protein levels (1,363 mg/L), while glucose level was highly decreased (12 mg/dL). Neuropathologic cytologic analysis of the CSF demonstrated malignant epithelial cells (expressing the epithelial membrane antigen) in terms of a meningeosis neoplastica. MRI of the brain on day 4 revealed several infratentorial and supratentorial lesions (figure, A). This was suggestive for cerebral metastasis of the esophageal carcinoma. CT scan of the chest and abdomen on day 6 showed partial atelectasis of the pulmonary right lower lobe and typical postoperative changes with reactively enlarged

mediastinal lymph nodes. There was evidence of neither COVID-19 pneumonia nor for local recurrence of the adenocarcinoma or further organ metastases.

Microbiological culture and a universal bacterial 16S-rDNA PCR were negative in the CSF sample from day 2. Because a nasopharyngeal swab was positive for SARS-CoV-2 RNA 1 day before admission to our hospital, further diagnostics for SARS-CoV-2 was initiated. Real-time RT-PCR analyses, using an in-house method targeting the SARS-CoV-2 E gene (screening assay) and the RdRp gene (confirmatory assay), were performed according to a previously published protocol.² On day 2, both screening and confirmatory assay were positive for SARS-CoV-2 RNA in the CSF (cycle threshold [ct] 19.5 and 21.6, respectively) and in the serum sample (ct 28.8 and 32.9, respectively), while PCR analysis of a nasopharyngeal swab was negative on day 5. Seroconversion was detected by an in-house fluorescence-activated cell sorting (FACS) antibody assay, using the SARS-CoV-2 full-length S protein, for immunoglobulin M (IgM) antibodies on day 2

Figure MRI and Fluorescence-Activated Cell Sorting (FACS)-Based Quantitative Measurement



(A) MRI of the brain on day 4. Upper row: axial T1-weighted MRI after IV contrast media application revealed 2 supratentorial and 2 infratentorial parenchymal lesions with slight perifocal edema (red circles). With regard to the adenocarcinoma of the esophagus, these lesions were suggestive for cerebral metastasis of the esophageal carcinoma. Lower row (left and middle): axial T1-weighted MRI after IV contrast media application demonstrated slight generalized atrophy of the brain with subsequent slightly enlarged sulci and ventricles without clear signs of a hydrocephalus referring in particular to the temporal horns. Lower row (right): sagittal T1-weighted MRI after IV contrast media application demonstrated infratentorial leptomeningeal thickening and increased contrast enhancement (yellow circles). (B) FACS-based quantitative measurement of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)-specific immunoglobulin G (IgG) antibodies in serum and CSF. Seroconversion for SARS-CoV-2 IgG was detected on day 6. The SARS-CoV-2 IgG CSF/serum ratio for paired CSF/serum samples on day 23 and day 26 was 140×10^{-3} and 90×10^{-3} , respectively. Based on total IgG and albumin values for CSF and serum, the SARS-CoV-2 IgG specific antibody index (AI) was 5.1 for both pairs. This verified a SARS-CoV-2-specific intrathecal IgG antibody synthesis. < = under detection limit; n.d. = not determined.

and for IgG antibodies on day 6, verifying an acute SARS-CoV-2 infection.³ Furthermore, IgM (not shown) and IgG antibodies to SARS-CoV-2 were also detected in CSF from day 23 and 26 (figure, B). Clinically, there was a significant reduction of the headache on day 3. In follow-up PCR analyses, SARS-CoV-2 RNA was detected in low concentration in a CSF sample on day 20 (ct value 33.9), whereas the last 2 CSF and serum samples obtained on day 23 and day 26 were PCR-negative.

Determination of the SARS-CoV-2-specific IgG antibody indices (AIs) for these 2 CSF/serum pairs was performed by using the FACS antibody assay for quantitation of SARS-CoV-2 IgG antibodies and kinetic nephelometry for measurement of total IgG concentrations in CSF and serum. The SARS-CoV-2 IgG CSF/serum ratios were 140×10^{-3} and 90×10^{-3} on day 23 and day 26, respectively. In comparison, the total IgG quotients were only 27.7×10^{-3} and 17.6×10^{-3} for the 2 CSF/serum pairs. The SARS-CoV-2 IgG-specific AI, calculated by dividing the pathogen-specific IgG quotient by the total IgG quotient,⁴ was 5.1 for both pairs. In cases without pathogen-specific intrathecal antibody synthesis, the AI is expected to be 1.0 (range 0.5–1.5). Thus, the elevated AI of 5.1 in 2 CSF/serum pairs of this patient, 5 times higher than expected from IgG diffusion across the blood–brain barrier, demonstrated intrathecal antibody synthesis. This is an additional indicator for active or past SARS-CoV-2 replication in the CNS compartment.

Later in the course of disease, after elimination of the virus, intrathecal chemotherapy and radiotherapy were initiated due to progression of the cerebral metastases.

Discussion

Our report describes the first case of SARS-CoV-2 meningitis in a patient with meningeosis carcinomatosa. SARS-CoV-2 infection of the CNS was confirmed by detection of viral RNA in 2 independent CSF samples and SARS-CoV-2-specific intrathecal IgG antibody synthesis.

There is increasing awareness for the neurotropic potential of SARS-CoV-2, yet the exact mechanism for entry in the CNS is unknown. Infection via the olfactory epithelium and nerve or hematogenous infection by viremia are currently discussed routes for neuroinvasion.⁵ Meningeosis carcinomatosa might enhance the neuroinvasiveness of SARS-CoV-2 by increasing the permeability of the blood–brain barrier, or might allow spread of virus from the brain into hematogenous circulation. From a pathophysiologic perspective, the latter mechanism might be more likely in our patient. First, on day 2 the viral RNA concentration in CSF was more than 100-fold higher than in serum, verifying autochthonous virus replication within the CNS compartment. Second, detection of SARS-CoV-2 RNA in the blood is usually considered as a virologic

marker for severe pulmonary or systemic disease,⁶ but our patient did not present with severe respiratory or other extraneurologic symptoms.

SARS-CoV-2 infection of the CNS was also reported in an apparently immunocompetent patient.⁷ Therefore, it remains unclear whether neoplastic meningitis and immunosuppression are relevant risk factors for a neuroinvasive SARS-CoV-2 infection, or if both conditions appeared coincidentally in our patient without a pathophysiologic link. A limitation in the attribution of the patient's headache as a neurologic manifestation of COVID-19 is that this symptom could be also caused by the meningeosis carcinomatosa or cerebral metastasis. However, there was a significant improvement of the headache on day 3, which correlated with the beginning of the adaptive immune response of the patient, since IgM antibody seroconversion was detected on day 2. In addition, neither an antiedematous nor antineoplastic therapy was administered at that time.

Our case demonstrates that in patients with neurologic symptoms, PCR diagnosis for SARS-CoV-2 RNA from both respiratory and CSF specimen is required. In the course of neurologic SARS-CoV-2 disease, viral RNA might be undetectable in respiratory samples, while SARS-CoV-2 RNA can be detected in CSF for longer periods, in our case for 18 days. In addition, the detection of seroconversion and quantitation of SARS-CoV-2-specific antibodies in CSF and serum can contribute to the diagnosis of COVID-19-associated neurologic involvement.

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Disclosure

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Appendix Authors

Name	Location	Contribution
Philipp A. Steininger, MD	University Hospital of Erlangen, Germany	Conceptualized case, wrote the manuscript, performed the molecular diagnostics, interpreted data
Frank Seifert, MD	University Hospital of Erlangen, Germany	Treated the patient, collection of patient samples, reviewed the final manuscript
Stefanie Balk, MD	University Hospital of Erlangen, Germany	Treated the patient, data acquisition, reviewed the final manuscript
Joji Kuramatsu, MD	University Hospital of Erlangen, Germany	Treated the patient, reviewed the final manuscript
Andreas E. Kremer, MD, PhD	University Hospital of Erlangen, Germany	Treated the patient, revised the manuscript, reviewed the final manuscript

Appendix (continued)

Name	Location	Contribution
Roland Coras, MD	University Hospital of Erlangen, Germany	Performed the neuropathology, reviewed the final manuscript
Tobias Engelhorn, MD	University Hospital of Erlangen, Germany	Interpreted the CT and MRI, reviewed the final manuscript
Clara Maier	University Hospital of Erlangen, Germany	Performed the antibody tests, created the figure, reviewed the final manuscript
Matthias Tenbusch	University Hospital of Erlangen, Germany	Performed the antibody tests, created the figure, reviewed the final manuscript
Klaus Korn, MD	University Hospital of Erlangen, Germany	Co-wrote the manuscript, assisted with expertise, data interpretation, and discussion, reviewed the final manuscript

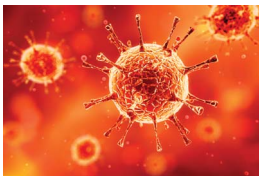
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Name	Location	Contribution
Armin Ensser, MD	University Hospital of Erlangen, Germany	Conceptualized case, wrote the manuscript, interpreted data

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